

Investigating RLS1 Localization and the Evolutionary Origins Of Cellular Differentiation

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ABSTRACT

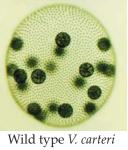
Cellular differentiation is a key attribute of all multicellular organisms, but little is known about the molecular mechanisms that drive its evolution. The goal of this work is to better understand a gene named RLS1 (regA-like sequence), that is believed to have been important for the evolution of cell types in the volvocine green algae. This family includes unicellular *Chlamydomonas reinhardtii*, which has no cell differentiation, and multicellular *Volvox carteri*. RLS1 is the closest C. reinhardtii homolog of RegA, a Volvox protein essential for cell differentiation; however, little is known about RLS1. Our immediate goal is to learn more about the accumulation and localization of the RLS1 protein during the *C. reinhardtii* life cycle by developing a construct that expresses mCherry-tagged RLS1 protein. We PCR-amplified a hygromycin-resistanceencoding gene fragment, which was subcloned into an *RLS1*-containing plasmid. An mCherry fragment was synthesized and will be subcloned to create the completed construct. That construct will be transformed into C. reinhardtii and western analysis will be used to identify transformants that express m-Cherry-RLS1. These transformants will be analyzed to determine how RLS1 accumulates during the *C. reinhardtii* life cycle, thus providing insights into its developmental function and the evolution of multicellularity.

BACKGROUND

Volvocine algae: *Volvox* and *Chlamydomonas*

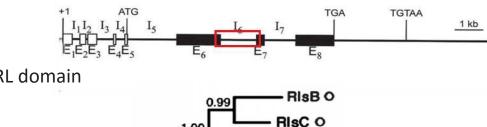


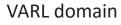
C. reinhardtii

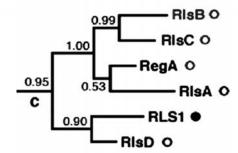




regA gene







- RegA contains a conserved region, the VARL (Volvocine Algal RegA) Like) domain that is found in homologs in both *V. carteri* and *C.* reinhardtii. RLS1 is the closest C. reinhardtii homolog of RegA.
- *RLS1* mRNA is expressed more when *C. reinhardtii* is light- or nutrientdeprived, which could mean that *RLS1* plays an important role in regulating photosynthesis.
- *RLS1* mRNA is present throughout the life cycle under normal growth conditions.

ACKNOWLEDGEMENTS

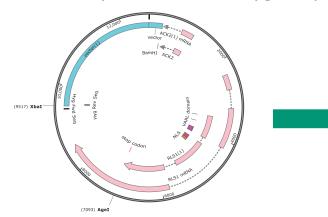
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OBJECTIVES

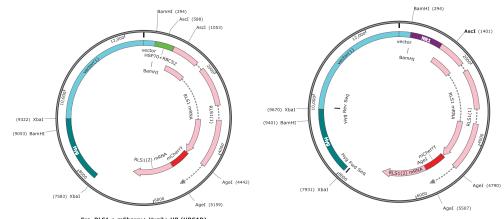
To better understand RLS1 and evolutionary origins of multicellularity in Volvocine algae by analyzing and manipulating protein expression.

METHODS

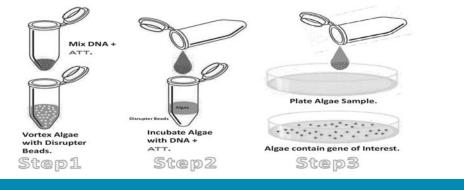
1. Create a construct that expresses mCherry-tagged RLS1 via native promoter, and hygromycin-resistance gene.



1. Create constructs with the HSP70A/RBCS (HR) promoter to overexpress RLS1 and NIT1 promoter to induce RLS1 expression in media that contains nitrate.

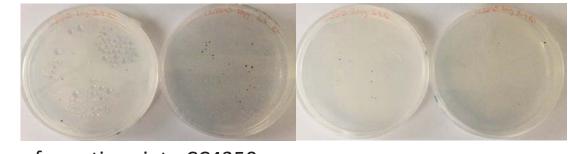


2. Transform all three constructs separately into the CC3395 and CC4350 strains of C. reinhardtii via glass bead transformation.

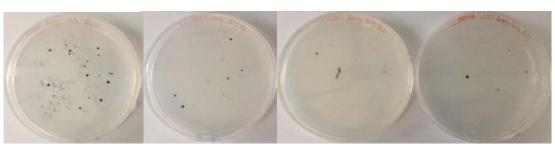


RESULTS

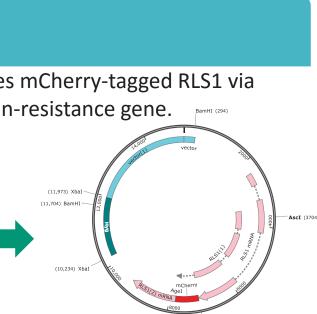
Transformations into CC3395 (selection on hygromycin)



Transformations into CC4350







RESULTS

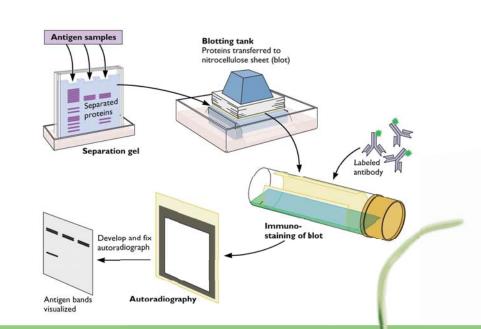
Strain	Transformation Type	Number of Colonies
CC3395	Control- pHyg	56
	1 ug of RLS1+mCherry+Hyg	32
	2 ug of RLS1+mCherry+Hyg	25
CC4350	Control- pHyg	52
	1 ug of RLS1+mCherry+Hyg	38
	2 ug of RLS1+mCherry+Hyg	16

CONCLUSIONS

- So far the mCherry+ Hyg+ RLS1 construct has been made and transformed into C. reinhardtii.
- From the transformations into CC4350 and CC3395, overall there were more transformants using less DNA (1ug).

FUTURE GOALS

- Complete the HR and NIT1 promoter constructs by inserting the hygromycin-resistant gene at the Xbal restriction site.
- Analyze transformants via confocal microscopy for red fluorescence (RLS1 protein).
- Perform Western blot analysis to analyze accumulation of the mCherry-RLS1 protein through out the life cycle.



REFERENCES

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